actuator 372 is positioned such that the valve cavity 374 is in fluid communication with the upper receptacle 320 prior to use. The device contains approximately 10 mg of autoclaved CM-111 3M Cosmetic Microspheres in the upper receptacle 320. Lower receptacle 324 contains a liquid detection reagent 365, which consists of approximately 0.6 milliliters of the luciferase/luciferin liquid reagent solution from a Clean-Trace surface ATP system. BARDAC 205M beads are made according to Preparative Example 5 of U.S. Patent Application No. 61/101,546, filed Sep. 30, 2008. Ten milliliters of sterile deionized water is added to the upper receptacle 320 of the unitary devices 300 immediately before use.

[0208] E. coli overnight cultures are prepared as described in Example 2. The bacterial culture is diluted to approximately  $10^6$  or  $10^5$  CFU/ml in Butterfield's buffer. One hundred microliters of the diluted suspension are pipetted directly into upper receptacle 320 of the unitary devices 300 to obtain a suspension of approximately 10<sup>5</sup> CFU or 10<sup>4</sup> CFU in ten milliliters, respectively. The cap 378 is used to close the housing 310 and the bacterial suspension is mixed with the microspheres (cell concentration agent 330) at room temperature and allowed to settle into the valve cavity 374. The cap 378 is removed and two BARDAC 205M beads (hydrogel 362) are dropped into the housing 310. Immediately after the beads settle into the valve cavity 374, the valve actuator 372 is turned to transfer the portion of the liquid sample in the valve cavity (containing the cell concentration agent 330 and the hydrogel 362) into the lower receptacle 324 containing the ATP detection reagents. The unitary device is immediately inserted into the reading chamber of a luminometer (for example, a NG Luminometer, UNG2) and RLU measurements are recorded at 10 sec interval using the Unplanned Testing mode of the UNG2 luminometer. RLU measurements are collected until the number of RLUs reaches a plateau. The data are downloaded using the software provided with the NG luminometer. The data will indicate that the microbial cells are concentrated by the microspheres, the cell extractant is released by the hydrogel, the cell extractant causes the release of ATP from the cells, and the ATP released from the cells is detected by the ATP detection system.

## Example 5

## Preparation of Detection Devices

[0209] Type I devices: For these detection devices, housings similar to the housing of FIG. 10A were constructed with the differences noted below. Reference numbers below refer to the corresponding parts in FIG. 10A. The upper parts 1012 and lower parts 1014 of the housing 1100 were obtained using the analogous components from 3M Clean-Trace<sup>TM</sup> surface ATP tests (obtained from 3M Company, Bridgend, UK). A collector 1067 with a frangible seal 1068 coupled thereto was press-fit into the upper portion of the lower part 1014; with the frangible seal 1068 facing the lower part 1014 of the housing 1100. The upper part 1012 was coupled to the lower part 1014 using a 2 cm section of 3:1 polyolefin dual wall adhesive lined heat shrink film obtained from buyheatshrink.com (part #\_HSC3A-050-cc, 1.5 cm in diameter) using a heat gun (Master Appliances Corp, Racine, Wis.).

**[0210]** For these detection devices, plungers similar to the plunger of FIG. **2**A were constructed. Reference numbers below refer to the corresponding parts in FIG. **2**A. The plunger (**250**) was assembled using a portion of the polyolefin plastic handle (**252**) from a 3M Clean-Trace<sup>TM</sup> surface ATP

test, a brass metal shaft (251) and an acetal piercing member 259. The handle 252 and piercing member 259 were attached to the ends of the brass shaft via threaded connections. The brass metal shaft was 11.5 cm long and 3.9 mm in diameter. A 6 mm, 6-23 thread was produced on each end of the shaft using a lathe. The piercing member 259 was fabricated from ½-inch (12.7 mm) acetal copolymer rod (part number 8497K211, obtained from McMASTER-CARR, Santa Fe Springs, Calif.) using a 10" Southbend lathe. An O ring (Buna N AS568A Dash Number 010 obtained from McMASTER-CARR) was used as the lower seal 256 and was attached to the plunger 250 approximately 11.5 mm above the piercing end 259. The plunger was surface-sterilized before each use.

[0211] Type II devices: These detection devices were assembled using a plunger similar to that shown and described in FIG. 5A with a tip similar to that shown in FIG. **6**A. The housing was constructed as described for the Type I devices. The tip of the plunger was fabricated from ½-inch (12.7 mm) acetal copolymer rod (part number 8497K211, obtained from McMASTER-CARR, Santa Fe Springs, Calif.) using a 10" Southbend lathe. The a duckbill one-way valve and a plastic retaining washer were press-fit into the recessed opening of the body of the tip of the plunger. The filter was made by machining a POREX filter (part number X6854 from Porex Corporation, Fairburn, Ga.) to the shape shown in FIG. 6A and dimensioning one end to press-fit into the recessed opening of the tip and hold the valve and retaining washer in place. The plunger was surface-sterilized before each use.

[0212] Type III devices: Detection devices similar to those shown in FIG. 10A were constructed with the differences noted below. Reference numbers below refer to the corresponding parts in FIG. 10A. The upper parts 1012 and lower parts 1014 of the housing 1100 were obtained using the analogous components from 3M Clean-Trace™ surface ATP tests (obtained from 3M Company, Bridgend, UK). A collector 1067 with a frangible seal 1068 coupled thereto was press-fit into the upper portion of the lower part 514; with the frangible seal 1068 facing the lower part 1014 of the housing 1100. The upper part 1012 was coupled to the lower part 1014 using a 2 cm section of 3:1 polyolefin dual wall adhesive lined heat shrink film obtained from buyheatshrink.com (part #\_HSC3A-050-cc, 1.5 cm in diameter) using a heat gun (Master Appliances Corp, Racine, Wis.).

[0213] The plunger (1050) was assembled using a portion of the polyolefin plastic handle (1052) from a 3M Clean-Trace<sup>TM</sup> surface ATP test, a brass metal shaft (1051) and tip 1090. The handle 1052 and tip 1090 were attached to the ends of the brass shaft via threaded connections. The brass metal shaft was 11.5 cm long and 3.9 mm in diameter. A 6 mm, 6-23thread was produced on each end of the shaft using a lathe. The tip 1090 was fabricated from ½-inch (12.7 mm) acetal copolymer rod (part number 8497K211, obtained from McMASTER-CARR, Santa Fe Springs, Calif.) using a 10"Southbend lathe. An O ring 1086 was attached to the tip 1090. The tip was machined to include a retaining member 1087, as shown in FIG. 10. A scraper was constructed by die-cutting a piece of 1 mm-thick polyurethane rubber and slipping it into the retaining member 1087. The outer diameter of the scraper 1086 was dimensioned to provide a tight fit with the inside of the housing 1010. The plunger was surfacesterilized before each use.